

Phytochemical evaluation and anticancer activity of *Ocimum sanctum* L. - A review

Ganesan Venkatachalam¹, Amutha Muthusamy^{2*}

ABSTRACT

Aim: The purpose of the paper is to review especially the phytochemical evaluation and anticancer activity of Tulsi. **Methods:** Several publications and books were electronically searched in Google using the keywords "Tulsi as a medicinal plant," "Tulsi and its anticancer activity," and "Anticancer activity of *Ocimum sanctum*. The search was restricted to books and articles related to anticancer activity of Tulsi from the year 1972 to 2017 by encountering the title and abstracts, and further short listing articles for full content. **Conclusion:** The present review revealed that *Ocimum sanctum* possesses an extensive anticancer efficacy still limited because of the lack of clinical trials on humans.

KEY WORDS: Anticancer activity, Ocimum sanctum, Tulsi

INTRODUCTION

Since 1000 years, human beings have been using medicinal plants for treating various diseases. Traditional medicines are, however, preferred by communities who cannot incurpharmaceutical products for their physical and psychological requirements. In several developing countries, especially in rural areas, people depend on traditional medicines for their primary health care. These are safer and cheaper than the synthetic medicines.^[1] Medicinal plants have been playing an important role all through human history. Medicinal plants have phytoconstituents that possess biological activity. Over 12000, such compounds have been isolated from various medicinal plants so far.^[2] More than 120 active compounds have been isolated from different medicinal plants, which are being used as herbal medicines. Nearly 80% of resemblance is shown between their modern therapeutic use and traditional. Traditional medicinal practices such as Siddha, Ayurveda, and Unani are studied for their chemical and pharmacological potential.[3]

Cancer is the second leading cause of human death. The American Cancer Society assessed that 1,665,540 new instances of tumor were required to be analyzed in 2014.

Access this article online		
Website: jprsolutions.info	ISSN: 0974-6943	

However, extraordinary gatherings of medications work in various approaches to battle growth cells and psychologist tumors, these days, herbs are utilized for malignancy cure.^[4,5] Chemotherapy may be used alone for certain types of cancer or in conjunction with other therapy such as radiation or surgery.^[6] Recent researches concentrate in developing suitable chemotherapy consistent with their new exploration in cell biology for the treatment of cancer with less or no toxic effect.^[7,8]

Ocimum sanctum L. (also known as Tulsi) has been used for several decades in Ayurveda for its assorted healing potential. Tulsi, the Queen of herbs, the unbelievable "Unique one" of India, is one of the holiest and most valued of the numerous mending and sound giving herbs of the orient. The hallowed basil, Tulsi, is prestigious^[9] for its religious and profound sacredness and for its critical part in the customary Ayurvedic and Unani arrangement of all-encompassing well-being and natural prescription of the East. It is specified by Charaka in the Charaka Samhita, an Ayurvedic content. Tulsi is thought to be an adaptogen, adjusting diverse procedures in the body, and supportive for adjusting to pressure. Set apart by its solid smell and astringent taste, it is viewed in Ayurveda as a sort of "remedy of life" and accepted to increase lifespan. Tulsi extracts are utilized as a part of Ayurvedic solutions for normal colds, cerebral pains, stomach issue, aggravation, coronary illness, and intestinal sickness. In general, O. sanctum L. is taken in different forms, as natural

¹Department of Chemistry, Nachimuthu Polytechnic College, Pollachi, Tamil Nadu, India, ²Department of Chemistry, Nallamuthu Gounder Mahalingam College, Pollachi, Tamil Nadu, India

*Corresponding author: Amutha Muthusamy, Department of Chemistry, Nallamuthu Gounder Mahalingam College, Pollachi, Tamil Nadu, India. E-mail: amutha@mail.com

Received on: 25-03-2018; Revised on: 21-04-2018; Accepted on: 26-05-2018

Journal of Pharmacy Research | Vol 12 • Issue 7 • 2018

tea, dried powder, or fresh leaf. For thousands of years, the dried leaves of Tulsi have been blended with stored grains to repel insects.^[10]

Family Description

- Kingdom: Plantae
- (Unranked) Angiosperms
- (Unranked) Eudicots
- (Unranked) Asterids
- Order: Lamiales
- Family: Lamiaceae
- Genus: Ocimum
- Species: *O. tenuiflorum*
- Binomial name: Ocimum tenuiflorum or O. sanctum L.

O. sanctum L. (Tulsi) is an erect, much-branched subshrub 30–60 cm tall, with simple opposite green or purple leaves that are strongly scented and hairy stems. Leaves have petiole and are ovate, up to 5 cm long, usually somewhat toothed. Flowers are purplish in elongate racemes in close whorls. Tulsi is native throughout the world tropics and widespread as a cultivated plant and an escaped weed. It is mainly cultivated for religious and medicinal purposes and for its essential oil. Tulsi is an important symbol in many Hindu religious traditions, which link the plant with Goddess figure. The name Sanskrit meaning of "Tulsi" is "the incomparable one."

The genus *Ocimum* is highly variable and possesses wide genetic diversity at intra- and inter-species levels. Nine species of *Ocimum*, namely *Ocimum tenuiflorum* L., *Ocimum basilicum* L., *Ocimum gratissimum* L., *Ocimum kilimandscharicum*, *Ocimum micranthum* L., *Ocimum campechianum* L., *Ocimum americanum* L., *Ocimum minimum* L., and *Ocimum citriodorum* L., are found in India, three of which (*O. americanum* L., *O. minimum* L., and *O. citriodorum* L.) are exotic.^[11] It is difficult to distinguish all these species on the basis of leaf morphology alone.

Phytochemical Constituents

Tulsi contains many nutrients and other biologically active compounds. The proportions of the phytoconstituents may vary significantly among strains and even among plants within the same field. The nutritional and pharmacological properties of Tulsi may due to the synergistic interactions of many different active phytochemicals. Consequently, the overall effects of Tulsi cannot be completely duplicated with extracts and isolated compounds. This may be due to the inherent botanical and biochemical complexity of Tulsi; institutionalization has, up until this point, evaded the present day science.

The volatile $oil^{[12]}$ obtained from the leaves of *O. sanctum* L. contains the phytochemicals shown in Table 1.

Seed oil^[12] has some fatty acids and sitosterols. Green leaves contain eugenol and methyl eugenol,^[16,17] anthocyanins, and certain levels of sugars, which are composed of xylose and polysaccharides present in seed mucilage. The leaves and stem contain various bioactive compounds such as flavonoids, saponins, tannins, and triterpenoids.^[14] The essential oil from leaves contains α -thujene, benzene, (Z)-3-hexanol, octane, nonane, ethyl 2-methyl butyrate, toluene, α-pinene, β-pinene, citronellal, sabinene, camphene, myrcene, dimethylbenzene, ethylbenzene, 1,8-cineole, limonene, p-cymene, Cis-βocimene, trans-β-ocimene, terpinolene, butyl-benzene, allo-ocimene, trans-linalool oxide, α-cubebene, geraniol, γ -terpene, α -copaene, β -cubebene, eugenol, linalool, α -guaiene, α -amorphene, β -bourbonene, β-caryophyllene, methyl eugenol, β-elemene, β-farnesene, (E)-cinnamyl acetate, isocaryophyllene, γ -humulene, 4, 11-selinadiene, borneol, iso-eugenol, myrtenylformat, α -humulene, α -terpineol, α -salilene, α -muurolene, β -salilene, δ -cadinene, calamenene, isoborneol, germacrene-D, iedol, humulene oxide, caryophyllene oxide, cuparene, geraneol, α -guaiol, τ -cadinol, nerolidol, α -bisabolol, cis-sesquisabinene hydrate, elemol, (EZ)-farnesol, tetradecanal, and 14-hydroxy-α-humulene selin-11-en-4-α-ol.^[18-25]

The alcoholic extract of the aerial parts of *O. sanctum* L. contains ursolic acid, rosmarinic acid, luteolin, apigenin, luteolin-7- O-glucuronide, isorientin, molludistin, apignin-7-O-glucuronide, stigmasterol, gallic acid, gallic acid methyl ester, β -stigmasterol triacontanol ferulate, orientin, vicenin-2, isovitexin, aesculectin, vitexin, aesculin, galuteolin, gallic acid ethyl ester, circineol, vanillin, chlorogenic acid, protocatechuic acid, phenylpropane glucosides-1, and phenylpropane glucosides-2.^[26-32]

The leaves contain carotene and ascorbic acid. The oil extracted from the seed of *O. sanctum* L. is called fixed oil which contains the following fatty acids stearic acid, palmitic acid, linoleic acid, oleic acid, sitosterol, linolenic acid hexourenicacid, linolenodilinolin, and dilinolenolinolins.^[33,34] The mineral content per 100 g of the whole plant contains carotene (2.5 mg), Vitamin C (83 mg), calcium - 3.15%, chromium - 2.9 μ g, phosphorous - 0.34%, zinc - 0.15 μ g, vanadium - 0.54 μ g, iron - 2.32 μ g, nickel - 0.73 μ g, copper - 0.4 μ g, and insoluble oxalate.^[35]

GC–MS evaluation of methanolic extract of leaves showed the presence of eugenol, methyl isoeugenol, and caryophyllene.^[36] Devendran *et al.* have identified the presence of 10 bioactive compounds in the hydroalcoholic extract of *O. sanctum* L. leaves by GC–MS analysis^[37] is shown in Table 2.

Anticancer Activity of O. sanctum L.

O. sanctum Linn. is a rich source of phytoconstituents and possesses an outstanding role in medicine. These

phytochemicals are not essential for its survival but are of substantial importance to human community to carry out various protective functions in human body.

Table 1: Phytoconstituents isolated from various parts of Ocimum sanctum L

Name of the compound	IUPAC naming	Structure
Eugenol ^[12]	1-hydroxy-2-methoxy-4-allylbenzene	
Ursolic acid ^[13]	2,3,4,5,6,6a, 7,8,8a, 10,11,12,13,14b-tetradecahydro-1H-picene-4a-carboxylic acid	
Carvacrol ^[13]	5-isopropyl-2-methylphenol	
Linalool ^[13]	3,7-dimethylocta-1,6-dien-3-ol	
Caryophyllene ^[13]	4,11,11-trimethyl-8-methylene-bicyclo[7.2.0]undec-4-ene	
Methyl chavicol ^[13] also called estragole	1-allyl-4-methoxybenzene	
Rosmarinic acid ^[14]	s(2 <i>R</i>)-2-[[(2 <i>E</i>)-3-(3,4-Dihydroxyphenyl)-1-oxo-2-propenyl]] oxy]-3-(3,4-dihydroxyphenyl) propanoic acid	
Apigenin ^[14]	5,7-dihydroxy-2-(4-hydroxyphenyl)-4H-1-benzopyran-4-one	
Cirsimaritin ^[14]	5,4'-dihydroxy-6,7-dimethoxyflavone	

(Contd...)

Table 1: (C	ontinued)
-------------	-----------

Name of the compound	IUPAC naming	Structure
Isothymusin ^[14]	6,7-dimethoxy-5,8,4'-trihydroxyflavone	
Isothymonin ^[14]	5,8,4'-Trihydroxy-6,7,3'-trimethoxyflavone	
Orientin ^[15]	8-C-beta-glucopyranosyl-3',4',5,7-tetrahydroxyflav-2-en-3-one	
Vicenin ^{sss}	6-C-beta-D-xylopyranosyl-8-C-beta-D-glucopyranosyl apigenin	

O. sanctum has been shown to possess anti-fatigue activity,^[38] adaptogenic property,^[39] antimicrobial activity,^[40,44] anticonvulsant activity,^[45,46] antifertility activity,^[47] antidiabetic activity,^[48] radioprotective activity,^[49,50] anti-inflammatory activity,^[51,54] cardioprotective activity,^[55,56] immunomodulatory activity,^[57] hepatoprotective activity,^[59,60] anticarcinogenic activity,^[61,62] mosquito repellent activity,^[63] and analgesic activity.^[64]

O. sanctum has been shown to possess an excellent anticancer activity.[65] Detoxification of mutagens and cancer-causing agents which is accomplished by enzymes such as cytochrome P450, glutathione-S-transferase, and cytochrome b5 and aryl hydrocarbon hydroxylase is regulated by the alcoholic leaf extract of O. sanctum. The anticancer efficacy of Tulsi against human fibrosarcoma cells culture has been reported; the results revealed that the alcoholic leaf extract induced cytotoxicity at the concentration of 50 mg/mL and above. The microscopical examination has shown the cells with contracted cytoplasm and shrunken nuclei. Agarose gel electrophoresis study has shown the DNA fragmentation.^[66] Various types of mutagen have been attempted for evaluating the anticarcinogenic ability in the experimental animals induced by O. sanctum leaves when fed to experimental rats for 10 weeks with 600 mg/g diet, promisingly reduced the 3,4-benzo (a) pyrene [B (a) P] and 3'-methyl-4-dimethylaminoazobenzene (3'MeDAB)-induced squamous cell carcinoma and hematoma incidences.^[67]

Prashar et al. in their study suggested that Tulsi leaf extract obstructs or overcomes biochemical issues accompany with chemical carcinogenesis by hampering metabolic stimulation of the procarcinogen to carcinogen. Primary cultures of rat hepatocytes were treated with 0-500 µg of O. sanctum L. extract for 24 h and then with 7,12-dimethaylbenz[a] anthracene (DMBA, 10 or 50 µg) for 18 h. Cells were then harvested and their DNA was isolated and analyzed by 32 p postlabeling. A significant reduction in the levels of DMBA/DNA adducts was observed in all cultures pretreated with O. sanctum L. extract. Hepatocytes that were treated with the highest dose of extract (500 µg) showed a maximum reduction of 93% in the mean values of DMBA/DNA adducts. This suggests the inhibition of metabolic activation of carcinogen.^[68] Papilloma genesis induced by DMBA (7,12-dimethylbenz[a]anthracene) in mice has shown promising reduction in tumor size on topical application of Tulsi leaf extract. Banerjee et al. in their study reported that the application of O. sanctum leaf extracts in the form of paste has shown significant prevention of DMBA-induced buccal pouch carcinogens.^[69]

O. sanctum ethanol leaf extract (70%) significantly reduces the incidence of cancer induced by *N*-methyl-*N'*-nitro-*N*-nitrosoguanidine (MNNG), a nitroso compound extensively used as an experimental gastric mutagen. MNNG is an efficient carcinogen and induces destruction of gastric mucosa which initiates stomach carcinogenesis. Intragastric administration

Name of the identified compound	Molecular formula
Eugenol	$C_{10}H_{12}O_{2}$
Caryophyllene	$C_{10}H_{12}O_{2}$ $C_{15}H_{24}$
Cyclohexane, 1,2,4-triethenyl-	C ₁₀ H ₁₀
Pentanedinitrile, 2-methyl-	$C_{6}^{12}H_{8}N_{2}^{18}$
10-Heptadecen-8-ynoic acid, methyl ester, (E)-	$C_{1}^{\dagger}H_{20}O_{2}$
Benzene methanamine, N, N-a, 4-tetramethyl-	$C_{11}H_{17}N^2$
Cyclopentane, cyclopropylidene-	$C_{s}^{1}H_{12}^{17}$
Z, Z-4,16-Octadecadien-1-ol acetate	$C_{20}^{\circ}H_{36}^{2}O_{2}$
3',8,8'-trimethoxy-3-piperidyl-2,2'-binaphthalene1,1',4,4'-tetrone	$C_{28}^{20}H_{25}^{30}NO_{7}$
Octadecane, 1,1-dimethoxy-	$C_{20}^{20}H_{42}^{23}O_2$

Table 2: GC-MS identified compounds from various extracts of Ocimum sanctum. L

of MNNG causes increased cell proliferation and angiogenesis with evasion of programmed cell death and leading to well-differentiated squamous cell carcinomas. Oral intake of *O. sanctum* has been shown to reduce these activities where Tulsi extract controls the critical molecules engage in invasion apoptosis, angiogenesis, and cell proliferation. A considerable decrease in the levels of cytokeratin, vascular endothelial growth factor (angiogenesis), antiapoptotic protein Bcl-2 glutathione-S-transferase pi (key proteins involved in proliferation), CK (infiltration), proliferating cell nuclear antigen with contemporary increase in cytochrome *c*, proapoptotic proteins Bax, and caspase 3 was disclosed.^[70]

Tulsi curtails the expression of γ -glutamyl transpeptidase, a marker of tumor sequence, and glutathione-S-transferase-P, which is increased in chemically induced hepatic tumors. The heat shock protein, which is distorted at the time of carcinogenesis, has also shown a decrease in its concentration.^[71] Tulsi extract reduced the efficacy of ornithine decarboxylase, an enzyme entangled in the regulation of cell proliferation and augmentation of cancer. There was also a contemporary reduction in the Phase I enzymes and lipid peroxidation implying that O. sanctum hinders the activity of carcinogen caused cytochrome P-450 - reliant enzymes and that this causes a decrease in the evolution of ultimate carcinogenic moiety.^[72] The anticancer efficacy^[73] of seed oil of Tulsi was examined against subsequently injected 20-methylcholanthrene-induced fibrosarcoma tumors in the thigh region of Swiss albino mice. Augmentation of 100 µL/kg body weight (maximum tolerated dose) of the oil significantly reduced 20-methaylcholathrene-induced tumor prevalence and tumor volume. The improved survival rate and delayed tumor occurrence were noted in seed oil supplemented mice.

CONCLUSION

Cancer keeps on being an overall killer in spite of great advances made in current system of medication amid the past decades. As indicated by late measurable information, malignancy is the second most basic reason for death after coronary illness.^[74] Cancer is a hyperproliferative ailment which causes invasion, proliferation, angiogenesis, dysregulation of apoptosis, and metastasis.^[75] Oncologists have noticed that advanced stage of cancers is impossible to treat. The reason behind the initiation, development, and evolution of cancer is not clearly known. Some oncologists believe that cancer is not a disease, anaerobic cell growth that ingests the carcinogens which kills patients. Oncologist Professor, Dr. Jones, declares "My studies have proven conclusively that cancer patient who refuses chemotherapy and radiation actually live up to 4 times longer than treated cases including untreated breast cancer cases."[76] There is a great scope and potential for the plantderived compounds in the fight to control or delay the evolution of the carcinogenic process without any side effects.

Tulsi, a traditional medicinal herb, considered as sacred plant, grown all over India, has been used as adaptogen helping the body and mind to adopt and cope with a wide range of emotional, physical, and stress. O. sanctum Linn. is a rich source of secondary metabolites which has a significant physiological effects in humans and is used as medicine. From the results of various studies, and the isolated and identified secondary metabolites evident that the whole plant remarkably controls and reduces many diseases. Anticancer property of Tulsi may be due to the synergistic effect of various phytoconstituents present in it. Daily usage of Tulsi leaves helps in controlling various diseases and disease-associated pathogens. By this review, it is clear that so many studies have been accomplished in the field of medicine with animal models to prove its efficacy in the treatment of cancer. Further, clinical trials need to be carried out on humans to ascertain the real effects of this "holy basil."

REFERENCES

- Ammara H, Salma R, Farah D, Shahid M. Antimicrobial activity of some plant extracts having hepato-protective effects. J Med Plant Res 2009;3:20-3.
- 2. Gupta R, Gabrielsen B, Ferguson SM. Nature's medicines: Traditional knowledge and intellectual property management.

Case studies from the national institutes of health (NIH), USA. Curr Drug Discov Technol 2005;2:203-19.

- Sandhu DS, Heinrich M. The use of health foods, spices and other botanicals in the Sikh community in London. Phytother Res 2005;19:633-42.
- Madhuri S, Pandey G. Some dietary agricultural plants with anticancer properties. Plant Arch 2008;8:13-6.
- Agarwal N, Majee C, Chakraborthy GS. Natural herbs as anticancer drugs. Int J PharmTech Res 2012;4:142-53.
- Reichardt P, Lindner T, Pink D, Thuss-Patience PC, Kretzschmar A, Dörken B, *et al.* Chemotherapy in alveolar soft part sarcomas. What do we know? Eur J Cancer 2003;39:1511-6.
- Yin SY, Wei WC, Jian FY, Yang NS. Therapeutic applications of herbal medicines for cancer patients. J Evid Based Complementary Altern Med 2013;2013:15.
- Pandey G, Madhuri S. Medicinal plants: Better remedy for neoplasm. Indian Drugs 2006;43:869-74.
- Warrier PK. In: Longman O, editor. Indian Medicinal Plants. New Delhi: CBS Publication; 1995. p. 168.
- Biswas NP, Biswas AK. Evaluation of some leaf dusts as grain protectant against rice weevil *Sitophilus oryzae* (Linn.). Environ Ecol 2005;23:485-8.
- 11. Willis JC, editor. A Dictionary of the Flowering Plants and Ferns. Cambridge: The University Press; 1919.
- Kelm MA, Nair MG, Stasburg GM, DeWitt DL. Antioxidant and cyclooxygenase inhibitiory phenolic compounds from *Ocimum sanctum* Linn. Phytomedicine 2000;7:7-13.
- 13. Shishodia S, Majumdar S, Banerjee S, Aggarwal BB. Urosolic acidinhibits nuclear factor-kappa B activation induced by carcinogenic agents through suppression of Ikappa Balpha kinase and p65 phosphorylation: Correlation with downregulation of cyclooxygenase 2, matrix metalloproteinase 9, and cyclin D1. Cancer Res 2003;63:4375-83.
- Jaggi RK, Madaan R, Singh B. Anticonvulsant potential of holy basil, *Ocimum sanctum* Linn., and its cultures. Indian J Exp Biol 2003;41:1329-33.
- Uma Devi P, Ganasoundari A, Vrinda B, Srinivasan KK, Unnikrishnan MK. Radiation protection by the *Ocimum* flavonoids orientin and vicenin: Mechanisms of action. Radiat Res 2000;154:455-60.
- Patil KS, Bhardwaj LK, Juvatkar PV, Shukla VK, Manvi FK. Plant products potential as anti-angiogenic and in cancer management. Int J Res Ayureda Pharm 2010;1:339-49.
- Kuhn M, Winston D. Winston and Kuhn's Herbal Therapy and Supplements: A Scientific and Traditional Approach. USA: Lippincott Williams and Wilkins; 2007. p. 260.
- Phillip MP, Damodaran NP. Chemo-types of *Ocimum sanctum* Linn. Indian Perfumer 1985;29:49-56.
- Kothari SK, Bhattacharya AK, Ramesh S. Essential oil yield and quality of methyl eugenol rich *Ocimum tenuiflorum* L.f. (syn *Ocimum sanctum* L.) grown in south India as influenced by method of harvest. J Chromatogr A 2004;1054:67-72.
- Lawrence BM, Hogg JW, Terhune SJ, Pichitakul N. Essential oils and their constituents. IX. The oils of *Ocimum sanctum* and *Ocimum basilicum* from Thailand. Flav Industry 1972;9:47-9.
- Brophy JJ, Goldsack RJ, Clarkson JR. The essential oil of Ocimum tenuiflorum L. (Lamiaceae) growing in Northern Australia. J Essent Oil Res 1993;5:459-46.
- Machado MI, Silva MG, Matos FJ, Craverio AA, Alencar WJ. Volatile constituents from leaves and inflorescence oil of *Ocimum tenuiflorum* L.f. (syn *Ocimum sanctum* L) grown in Northeastern Brazil. J Essent Oil Res 1999;11:324-6.
- Vina A, Murillo E. Essential oil composition from twelve varieties of Basil (*Ocimum* spp) grown in Colombia. J Braz Chem Soc 2003;14:744-9.
- Dey BB, Choudhury MA. Essential oil of *Ocimum sanctum* L. and its antimicrobial activity. Indian Perfumer 1984;28:82-7.
- Dey BB, Choudhary MA. Effect of plant development stage and some micronutrients on eugenol content in *O. sanctum* L. determination of eugenol by folin-ciocalteu reagent. Indian Perfumer 1980;24:199-203.
- 26. Sukari MA, Rahmani M, Lee GB. Constituents of stem barks of

Ocimum sanctum. Fitoterapia 1995;64:552-3.

- 27. Nguyen H, Lemberkovics E, Tarr K, Mathe IJ, Petri G. A comparative study on formation of flavonoid, tannin, polyphenol contents in ontogenesis of *Ocimum basilicum* L. Acta Agron Hung 1993;42:41-50. British Journal of Pharmaceutical Research 2013;3(2):273-292.
- Skaltsa H, Philianos S, Singh M. Phytochemical study of the leaves of *Ocimum sanctum*. Fitoterapia 1987;8:286.
- Nörr H, Wagner H. New constituents from Ocimum sanctum. Planta Med 1992;58:574.
- Nair AG, Gunasegaran R. Chemical investigation of certain South Indian plants. Indian J Chem 1982;21B:979-80.
- Skaltsa H, Tzakou O, Singh M. Polyphenols of Ocimum sanctum from suriname. Pharmaceut Bio 1999;37:92-4.
- Kicel A, Kurowska A, Kalemba D. Composition of the essential oil of *Ocimum sanctum* L. grown in Poland during vegetation. J Essent Oil Res 2005;17:217-19.
- Nadkarni GB, Patwardhan VA. Fatty oil from the seeds of Ocimum sanctum Linn. (Tulsi). Cur Sci 1952;91:68-9.
- Singh S, Majumdar DK, Yadav MR. Chemical and pharmacological studies of *Ocimum sanctum* fixed oil. Indian J Exp Biol 1996;34:1212-5.
- Naredhirakannan RT, Subramanian S, Kandaswamy M. Mineral content of some medicinal plants used in the treatment of diabetes mellitus. Biol Trac Elem Res 2005;105:109-16.
- Balasubramanian S, Ganesh D, Reddy PS, Narayanan VV. GC-MS analysis of phytocomponents in the methanolic extract of *Ocimum Sanctum* (Tulsi). Asian J Pharm Anal Med Chem 2014;2:71-5.
- Devendran G, Balasubramanian U. Qualitative phytochemical screening and GC-MS analysis of *Ocimum sanctum* L. leaves. Asian J Plant Sci 2011;1:44-8.
- Prasad MP, Khanum F. Antifatigue activity of ethanolic extract of *Ocimum sanctum* in rats. Res J Med Plant 2012;6:37-46.
- Gupta P, Yadav DK, Siripurapu KB, Palit G, Maurya R. Constituents of *ocimum sanctum* with antistress activity. J Nat Prod 2007;70:1410-6.
- Sagar A, Thakur I. Antibacterial activity of Ocimum sanctum (Linn.), Murraya koenigii (Linn.) spreng and Artemisia vulgaris (Linn.). Plant Arch 2012;12:377-81.
- Goyal P, Kaushik P. *In vitro* evaluation of antibacterial activity of various crude leaf extracts of Indian sacred plant, *Ocimum sanctum* L. Br Microbiol Res J 2011;1:70-8.
- Mishra P, Mishra S. Study of antibacterial activity of *O. sanctum* extracts against gram negative and gram positive bacteria. Am J Food Technol 2011;6:336-41.
- Singh S, Malhotra M, Majumdar DK. Antibacterial activity of Ocimum sanctum L. Fixed oil. Indian J Exp Biol 2005;43:835-7.
- 44. Geeta, Vasudevan DM, Kedlaya R, Deepa S, Ballal M. Activity of Ocimum sanctum (the traditional Indian medicinal plant) against the enteric pathogens. Indian J Med Sci 2001;55:434-8, 472.
- Sakina MR, Dandiya PC, Hamdard ME, Hameed A. Prelimnary psychopharmacological evaluation of *Ocimum sanctum* leaf extract. J Ethnopharmacol 1990;28:143-50.
- 46. Shiva MP. Inventory of Forestry Resources for Sustainable Management and Biodiversity Conservation. New Delhi: Indus Publishing Company; 1996.
- 47. Khan MR, Islam MA, Hossain MS, Asadujjaman M, Wahed MI, Rahman BM, *et al.* Antidiabetic effects of the different fractions of ethanolic extracts of *Ocimum sanctum* in normal and alloxan induced diabetic rats. J Sci Res 2010;2:158-68.
- Joseph LJ, Bhartiya US, Raut YS, Hawaldar RW, Nayak Y, Pawar YP, *et al.* Radioprotective effect of *Ocimum sanctum* and amifostine on the salivary gland of rats after therapeutic radioiodine exposure. Cancer Biother Radiopharm 2011;26:737-43.
- 49. Devi PU, Bisht KS, Vinitha M. A comparative study of radioprotection by *Ocimum* flavonoids and synthetic aminothiol protectors in the mouse. Br J Radiol 1998;71:782-4.
- Singh S. Comparative evaluation of antiinflammatory potential of fixed oil of different species of *Ocimum* and its possible mechanism of action. Indian J Exp Biol 1998;36:1028-31.

- Singh S, Taneja M, Majumdar DK. Biological activities of ocimum sanctum L. Fixed oil – an overview. Indian J Exp Biol 2007;45:403-12.
- Godhwani S, Godhwani JL, Vyas DS. Ocimum sanctum: An experimental study evaluating its anti-inflammatory, analgesic and antipyretic activity in animals. J Ethnopharmacol 1987;21:153-63.
- Sharma M, Kishore K, Gupta SK, Joshi S, Arya DS. Cardioprotective potential of *Ocimum sanctum* in isoproterenol induced myocardial infarction in rats. Mol Cell Biochem 2001;225:75-83.
- Panda VS, N SR. Evaluation of cardioprotective activity of Ginkgo biloba and Ocimum sanctum in rodents. Altern Med Rev 2009;14:161-71.
- 55. Mondal S, Varma S, Bamola VD, Naik SN, Mirdha BR, Padhi MM, *et al.* Double-blinded randomized controlled trial for immunomodulatory effects of tulsi (*Ocimum sanctum* linn.) leaf extract on healthy volunteers. J Ethnopharmacol 2011;136:452-6.
- Goel A, Singh DK, Bhatia AK. Effect of Ocimum sanctum extract on the induction of IFN-γ and IL-10 cytokines and their m-RNA expression. J Immunol Immunopathol 2010;12:29-41.
- Sujatha K, Srilatha CH, Anjaneyulu Y, Rao TS, Sreenivasulu D, Amaravathi P. Evaluation of hepatoprotective activity of *Ocimum sanctum* (Os) leaf extract on lead induced hepatic damage in wistar Albino rats. Invent Impact 2011;2011:Article ID: 290805.
- Akilavalli N, Radhika J, Brindha P. Hepatoprotective activity of *Ocimum sanctum* Linn. against lead induced toxicity in Albino rats. Asian J Pharm Clin Res 2011;4:84-7.
- 59. Nagaprashantha LD, Vatsyayan R, Singhal J, Fast S, Roby R, Awasthi S, *et al.* Anti-cancer effects of novel flavonoid vicenin-2 as a single agent and in synergistic combination with docetaxel in prostate cancer. Biochem Pharmacol 2011;82:1100-9.
- 60. Jha AK, Jha M, Kaur J. Ethanolic extracts of Ocimum sanctum, Azadirachta indica and Withania somnifera cause apoptosis in SiHa cells. Res J Pharm Biol Chem Sci 2012;3:557-62.
- Singh S, Mahour K, Prakash S. Evaluation of mosquito repellent efficacy of *Ocimum sanctum* plant extract. J Herb Med Toxicol 2009;3:87-90.
- 62. Singh S, Majumdar DK. Analgesic activity of *Ocimum* sanctum and its possible mechanism of action. Pharm Biol

1995;33:188-92.

- Devi PU. Radioprotective, anticarcinogenic and antioxidant properties of the Indian holy basil, *Ocimum sanctum* (Tulasi). Ind J Exp Biol 2000;39:185-90.
- Karthikeyan K, Ravichadran P, Govindasamy S. Chemopreventive effect of *Ocimum sanctum* on DMB-A induced hamster buccal pouch carcinogenesis. Oral Oncol 1999;35:11-29.
- Sporn MB, Suh N. Chemoprevention of cancer. Carcinogenesis 2000;21:525-30.
- 66. Prashar R, Kumar A, Hewer A, Cole KJ, Davis W, Phillips DH. Inhibition by and extract of *Ocimum sanctum* of DNAbinding activity of 7,12 dimethylbenz[a] anthracene in rat hepatocytes *in vitro*. Cancer Lett 1998;128:155-60.
- Banerjee S, Prashar R, Kumar A, Rao AR. Modulatory influence of alcoholic extract of *Ocimum* leaves on carcinogen induced metabolizing enzyme activities and reduced glutathione levels in mouse. Nutr Cancer 1996;25:205-17.
- Manikandan P, Murugan RS, Abbas H, Abraham SK, Nagini S. *Ocimum sanctum* Linn. (Holy Basil) ethanolic leaf extract protects against 7, 12 dimethylbenz(a) anthraceneinduced genotoxicity, oxidative stress, and imbalance in xenobioticmetabolizing enzymes. J Med Food 2007;10:495-502.
- Watson RR, Preedy VR. Bioactive Foods and Extracts. Cancer Treatment and Prevention. 1st ed. United States of America: CRS Press; 2011.
- Devi PU, Gonasoundari A, Vrinda B, Srinivasan KK, Unnikrishanan MK. Radiation protection by the *Ocimum* sanctum flavonoids orientin and vicenin: Mechanism of action. Radiat Res 2000;154:45560.
- Prakash J, Gupta SK. Chemopreventive activity of Ocimum sanctum seed oil. J Ethnopharmacol 2000;72:29-34.
- Jemal A, Siegel R, Ward E, Murray T, Xu J, Thun MJ, et al. Cancer statistics, 2007. CA Cancer J Clin 2007;57:43-66.
- Aggarwal BB, Ichikawa H, Garodia P, Weerasinghe P, Sethi G, Bhatt ID, *et al.* From traditional ayurvedic medicine to modern medicine: Identification of therapeutic targets for suppression of inflammation and cancer. Expert Opin Ther Targets 2006;10:87-118.
- Moritz A. Cancer is Not a Disease. 2nd ed. USA: Ener-Chi Wellness Press; 2008. p. 21-3.